



## Multiplex PCR screening of HBV, HIV and CMV.

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This protocol describes the PCR components and conditions used for the screening of three blood-borne pathogens through multiplexed nested endpoint PCR. This protocol allows for the identification of human immunodeficiency virus (HIV-1), hepatitis B virus (HBV) and human cytomegalovirus (CMV). It includes a KIR3DL2 framework gene inner control of larger size than target amplicons.

### Oligonucleotide primers used

Name	Target	PCR	Sequence	bp	%GC	Tm <sup>b</sup>	Position	Amplicon	Ref	
HIV-FO	HIV-1	1	5'-TAC-Agg-AgC-AgA-TgA-TAC-Ag-3'	20	45	50	141-161	294	1	
HIV-RO			5'-CCT-ggC-TTT-AAT-TTT-ACT-gg-3'	20	40	48	418-438			
HIV-FI		2	5'-ggA-AAC-CAA-AAA-TgA-TAg-gg-3'	20	40	48	221-241	130		
HIV-RI			5'-ATT-ATg-TTg-ACA-ggT-gTA-gg-3'	20	40	48	331-351			
HBV-FO	HBV	1	5'-CAC-CAT-gCA-ACT-TTT-TCA-CCT-CTg-C-3'	25	48	58	1810-1835	561	2	
HBV-RO			5'-TCT-gCg-Agg-CgA-ggg-AgT-TCT-3'	21	62	58	2375-2396		3	
HBV-FI		2	5'-AAg-CCT-CCA-AgC-TgT-gCC-TTg-g-3'	21	57	56	1866-1887	426	4	
HBV-RI			5'-gCA-ggA-ggA-gTg-CgA-ATC-CAC-AC-3'	23	61	61	2266-2289			
CMV-FO2	CMV	1	5'-gAA-TTC-gCg-CAT-gAT-CTC-3'	18	50	52	912-929	815	5	
CMV-RO2			5'-ggA-AAC-gTg-TCC-gTC-TT-3'	17	53	52	1711-1727			
CMV-FI2		2	5'-gCg-AgT-AAA-gTT-CCA-gTA-3'	18	44	49	968-985	719		4
CMV-RI2			5'-gTT-CTg-gCA-Agg-YA-3'	14	57	49	1675-1687			
F3DL2	3DL2	1	5'-TCA-TgC-TgT-ACA-AAg-AAg-ACA-gAA-g-3'	25	40	54	E3(45)	1713	7	
R3DL2			5'-ATg-ACT-gTC-TCT-CCT-gAT-TTC-Ag-3'	23	43	53	E4(113)			
3DL2FI		2	5'-TAC-AgA-TgT-Cgg-ggT-TCA-Cg-3'	20	55	57	E3	1500		
3DL2RI			5'-ggA-ggg-Aag-gTT-TTC-TgT-ggT-TTC-3'	24	50	58	E4			



## PCR components

First PCR		1 rx (μl)
dH <sub>2</sub> O	Cf	1.4
10x Buffer PCR	1X	1.25
MgCl <sub>2</sub> 50 mM	3.0 mM	0.75
4x dNTPs 10 mM	200 μM	0.25
Oligos HIV-ext. 10 μM	800 nM	1
Oligos HBV-ext 10 uM	400 nM	0.5
Oligos CMV-ext. 10 μM	800 nM	1
Oligos KIR3DL2-ext. 10 μM	200 nM	0.25
Taq 5 UI/μL	0.4 IU/μL	0.1
cDNA	-	6
	Vf	12.5

↓  
Run Multiplex 1

Second (nested) PCR		1 rx (μl)
dH <sub>2</sub> O	Cf	12.8
10x Buffer	1X	2.5
MgCl <sub>2</sub> 50 mM	2.0mM	1.0
4x dNTPs 10 mM	200 μM	0.5
Oligos HIV-int 10 μM	800 nM	2.0
Oligos HBV-int 10 uM	400 nM	1.0
Oligos CMV-int 10 μM	800 nM	2.0
Oligos KIR3DL2-int 10 μM (1/8)	50 nM	1.0
Taq 5 UI/μL	0.4 IU/μL	0.2
1:10 diluted 1 <sup>st</sup> PCR product	-	2.0
	Vf	25.0

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Run Multiplex 2

## PCR conditions

Total time: 1:58 hrs		Denaturing		Annealing		Extension	
Multiplex 1	Temperature	94°	94°	62°	72°	72°	4°
	Time	5 min	30 sec	30 sec	30 sec	5 min	5 min
		x30 cycles					

Total time: 1:58 hrs		Denaturing		Annealing		Extension	
Multiplex 2	Temperature	94°	94°	54°	72°	72°	4°
	Time	5 min	30 sec	30 sec	30 sec	5 min	5 min
		x30 cycles					

## Electrophoresis conditions

Add 5 μL of orange loading buffer to 25 μL of 2<sup>nd</sup> PCR, load 25μL into 1.5% agarose gel with at least 1.5 inch comb separation for 55 minutes at 130 VDC (5.2 V/cm).



## Notes

1. Clean work area with 70% Ethanol before and after preparing PCR master mix.
2. Prepare all PCRs in ice or cold-pack microtube racks to lower static evaporative rate and volume losses.
3. Vortex master mix before aliquoting into individual microtubes.
4. Prepare 1st and 2nd PCR mastermix in PCR enclosures. After cycling 1st PCR open tubes in post-PCR area ONLY and add 125  $\mu$ L of dH<sub>2</sub>O to dilute PCR product 1:10 for use in 2<sup>nd</sup> (nested) PCR.

## References

1. Albert, J. Simple, sensitive, and specific detection of Human Immunodeficiency Virus type 1 in clinical specimens by polymerase chain reaction with nested primers. *Journal of Clinical Microbiology* **28**, 1560 - 1564 (1990).
2. Aslam MA, Identification of Hepatitis B virus core mutants by PCR-RFLP in chronic Hepatitis B patients from Punjab, Pakistan. *Arch Virol* (2007).
3. Dia Sorin, Saluggia, Italia (comunicación personal).
4. CA García-Sepúlveda & SE Guerra-Palomares, Laboratorio de Biología Molecular, Facultad de Medicina UASLP México, 2008.
5. Chou SW. Analysis of interstrain variation in cytomegalovirus glycoprotein B sequences encoding neutralization-related epitopes. *J Infect Dis* (1991).
6. CA, García-Sepúlveda, Laboratorio de Genómica Viral y Humana, Facultad de Medicina UASLP, México, 2009.

## Revision history

- 1.0 Original document.
- 2.0 Changes to document format only.

